Product features

- Natural killer (NK) cells can be expanded from human peripheral blood mononuclear cells (PBMCs) without using feeder cells.
- NK cells can be expanded several hundred to several thousand-fold by 2 - 3 weeks of culture.
- One kit is sufficient to expand NK cells from 20 - 50 ml of whole blood.

<table>
<thead>
<tr>
<th>Kit name</th>
<th>Catalog No.</th>
<th>Amount</th>
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<tbody>
<tr>
<td>BINKIT®</td>
<td>N501-1</td>
<td>1 kit</td>
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<td></td>
<td>N501-2</td>
<td>2 kits</td>
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<td>4 kits</td>
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<td>N501-8</td>
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</table>

Kit components

One kit of BINKIT® includes:

- NK Cell Initial Flask (M) N104 1 flask (75 cm²)
- NK Cell Initial Medium N115a 45 ml
- NK Cell Initial Cocktail N115b 1.9 ml
- NK Cell Subculture Medium N201 1000 ml

Intended use

For research use only.
Not for use in diagnostic procedures.

Storage

Store at 2 - 10 °C. Protect from light.

Shelf life

One year after production or until expiration date.

Other supplies required

Ficoll-Paque (GE Healthcare, Sweden)
Sterile PBS
FBS or autologous plasma (It is desirable to be heat-inactivated at 56 °C for 30 minutes.)
Sterile conical centrifuge tubes
Sterile culture flasks

**Precautions**

NK Cell Initial Flask may carry condensation on the surface, which does not adversely affect the performance of the kit.

**Procedure overview**

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Preparation reagents

Preparing PBMCs
Washing NK Cell Initial Flask

Initial culture

Changing medium and sub-culturing

Harvesting CD3 CD56+ cells
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**Procedures**

**Preparing reagents**

NK Cell Initial Medium and NK Cell Subculture Medium should be supplemented with 5 % (v/v) of heat-inactivated FBS or autologous plasma.

**Preparing peripheral blood mononuclear cells (PBMCs)**

Isolate PBMCs from whole human blood by density gradient centrifugation using Ficoll-Paque.

**Washing NK Cell Initial Flasks**

Add 10 ml PBS to an NK Cell Initial Flask. Slant the flask to cover the entire surface with PBS. Aspirate the liquid completely from the flask. Care should be taken so as not to scratch the surface of the flask. Repeat the washing process two more times.
Culturing NK cells from PBMCs
Suspend the PBMCs in NK Cell Initial Medium at $1 \times 10^6$ cells/ml. Add 40 μl of NK Cell Initial Cocktail to 1 mL of the cell suspension. Transfer the cell suspension to the pre-washed NK Cell Initial Flask. Incubate under 5 % CO₂ at 37 °C for 3 days.

Changing medium on Day3
Transfer floating as well as adherent cells to a conical centrifuge tube and centrifugate at 200 x g for 8 minutes. Remove the supernatant and re-suspended the cells at $1 \times 10^6$ cells/ml in NK Cell Subculture Medium supplemented with 5 % (v/v) of heat-inactivated FBS or autologous plasma. The cell suspension is transferred to conventional culture flasks and is cultured under 5 % CO₂ at 37 °C.

Sub-culturing
Cells should be sub-cultured every 2 - 3 days by adding freshly NK Cell Subculture Medium at 0.8 x $10^6$ cells/ml.

Suggested culturing period
2 - 3 weeks.

Effects
CD3 CD56+ NK cells will be expanded several hundred to several thousand-fold in 2 - 3 weeks of culture, making more than 50 % of cultured cells to be CD3 CD56+ NK cells.

References
Xuewen Deng, Hiroshi Terunuma, Mie Nieda, Weihua Xiao, Andrew Nicol, Synergistic cytotoxicity of ex vivo expanded natural killer cells in combination with monoclonal antibody drugs against cancer cells, Int Immunopharmacol 14 (2012) 593-605

Xuewen Deng, Mie Nieda, Hiroshi Terunuma, Ex vivo expanded natural killer cells can possibly kill cancer stem cells, 18th ISCT Annual Meeting 2012
BINKIT® for NK cells expansion from PBMCs

Xuewen Deng, Hiroshi Terunuma, Mie Nieda, Cytotoxicity of expanded NK cells against cancer cells is enhanced by monoclonal antibody drugs, *19th ISCT Annual Meeting 2013*

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